

### 0271-5317(95)00042-9

### DETOXIFICATION METHODS OF AFLATOXINS. A REVIEW.

G. Piva<sup>1</sup>, FP, F. Galvano<sup>2\*</sup>, RD, A. Pietri<sup>1</sup>, AP, A. Piva<sup>3</sup>, RD.

<sup>1</sup>Istituto di Scienze degli Alimenti e della Nutrizione, Università Cattolica del Sacro Cuore, Piacenza.

<sup>2</sup>Istituto di Zootecnica, Università di Reggio Calabria.

<sup>3</sup>Istituto di Zootecnica e Nutrizione Animale, Università di Bologna.

#### **ABSTRACT**

Detoxification of aflatoxin contaminated foods and feeds is a current problem, as aflatoxins are highly carcinogenic and capable of passing unaltered through metabolic processes and accumulating in the tissues (seriously jeopardizing human and animal health). Although numerous detoxification methods have been tested, none seems able to fulfill the efficacy, safety, safeguarding of nutritional elements and costs requisites of a detoxification process. This paper critically reviews the main chemical detoxification methods and the latest approach to the problem using added sorbents capable of adsorbing aflatoxins.

KEY WORDS: Feeds, Foods, Aflatoxin, Toxicity, Detoxification, Sorbents.

### INTRODUCTION

Any decontamination process must be technically and economically feasible if it is to be applied practically. The FAO requirements for acceptable decontamination process (1) stipulate that the procedure must:

- 1. destroy, inactivate or remove aflatoxins;
- 2. not produce nor leave toxic and/or carcinogenic/mutagenic residues in the final products or in food products obtained from animals fed on decontaminated feeds;
- 3. not significantly alter the important technologic properties:

and ideally,

4. destroy fungal spores and mycelium that could proliferate and produce new toxins under favourable conditions.

Analogous requirements have been set down in France and U.S.A., however the F.D.A. (2) requires additional data on the impact of the process to the environment. Many detoxification methods of aflatoxin contaminated foodstuffs have been recommended and include: mechanical separation of contaminated seeds, heat treatment, extraction using solvents, detoxification using chemical agents and added sorbents. This paper reviews chemical treatments and added sorbents.

<sup>\*</sup>Address corrispondence and reprint request to: Dott. Fabio Galvano, Istituto di Zootecnica, Università di Reggio Calabria, P.zza S. Francesco di Sales, 7 - 89061, Gallina di Reggio Calabria, Italy.

# Treatment with Ammonia

Treatment with ammonia in the gaseous phase, in solution, or with substances capable of releasing it, achieved optimum results in detoxifying peanut, cotton and corn meals. It is legal in some of the North American states (Arizona, California, Georgia, Alabama) and French technology has been used to build some treatment plants in Senegal and more recently in France.

It has been widely demonstrated that the efficacy of detoxification with ammonia is positively correlated with the quantity used, reaction time, temperature and pressure levels and combination with formaldehyde (3). The mechanism of action induced by ammonia on the aflatoxin B<sub>1</sub> (AFB<sub>1</sub>) molecule has been studied extensively and seems to have been clarified. It was observed that the AFB<sub>1</sub> molecular structure is irreversibly altered if exposure to ammonia lasts long enough. In contrast, if exposure is not sufficiently protracted, the molecule can revert to its original state. In fact, some tests aimed at verifying carry-over in dairy cows fed ammonia treated feeds (4,5) revealed percentages of AFB<sub>1</sub> metabolite elimination in milk (i.e. aflatoxin M<sub>1</sub> or AFM<sub>1</sub>) between 10-20% compared with AFB<sub>1</sub> ingested. These percentages generally vary from 1-3%. This was explained by examining the detoxification reaction mechanism. The first step (reversible) consists of the opening of aflatoxin cyclic lactone (Fig. 1): in an acid environment (as in the stomach) the balance may shift towards the original products with consequent reformation of AFB<sub>1</sub>. In this case the animal would ingest a greater quantity of toxin compared with the quantity revealed analytically, explaining the higher percentage of AFM<sub>1</sub> in the milk.

The products which can trigger of the AFB<sub>1</sub>-ammonia reaction have diverse chemical characteristics. This variability may be determined by the presence of substances capable of interfering in the detoxification reaction and giving rise to not easily identified compounds in treated feed. Aflatoxin D<sub>1</sub> (AFD<sub>1</sub>) is one of the well known reaction products identified as a non fluorescent phenol with molecular weight (MW) 286 which reveals the absence of the lactone carbonylic functional group with respect to the AFB<sub>1</sub> molecule (6) (Figure 1).

FIG. 1 - Ammoniation of aflatoxin (8).

An analogous compound homogeneous for structure and molecular weight seems to form by reaction with sodium hydroxide at  $100^{\circ}$  C (7). In addition to the pathway leading to AFD<sub>1</sub> formation, the reaction can follow alternative pathways which can determine formation of other compounds. These include a series of non identified compounds with MW less than 200, a non fluorescent diffuran phenol with MW 206 obtained after formation of a  $\beta$ -keto acid (8), a compound

with MW 256 formed by  $AFD_1$  sublimation at temperatures between 220° and 340° C and a non identified compound with MW 236 produced after release of a  $CO_2$  molecule (9).

AFD<sub>1</sub>, AFM<sub>1</sub> and the chemical compound with MW 206 were biologically tested in order to evaluate the residual toxicity and mutagenic potential. Tests performed using salmonella microsomes, covalent binding index and chick-embryo development demonstrated that AFD<sub>1</sub> is much less toxic than AFB<sub>1</sub>. However, the data reported in the literature vary: 450 fold less (10); 280 fold less (11); 130 fold less (12) and from 2.000 to 20.000 times less (13). In contrast, AFM<sub>1</sub>'s toxicity and mutagenic potential are similar to those of the original molecule as shown by trials carried out on ducks (14), rats (15) and trout (16), whereas mutagenic potential seems to be reduced in rat kidneys (17). Studies conducted on broilers (18,19,20), laying hens (21) and rats (22,23) fed ammoniated AFB<sub>1</sub>-contamined feeds revealed no histologic alterations of target organs and improved performance parameters. A test on dairy cows (24) showed almost total disappearance of AFB<sub>1</sub> carry-over in milk (AFM<sub>1</sub> level was under 0.1  $\mu$ g/l).

The disadvantages of ammonia treatment are mainly related to the need to build special plants as ammonia corrodes metal and becomes explosive in the air at mixtures over 15% in volume. In addition, some effects on the chemical and qualitative characteristics of the feeds cannot be overlooked, these being the undesirable brown color of the treated feed, the increase in total nitrogen and non protein nitrogen together with a marked reduction in nitrogen solubility and diminished content of some amino acids (cystine, methionine and especially lysine), which is the main drawback of ammonia treatment.

#### Treatment with Sodium Bisulfite

Sodium bisulfite treatment is a valid AFB<sub>1</sub> detoxification strategy (25,26,27). Although it is less efficacious than ammonia detoxification it overcomes some of the typical disadvantages of ammonia methods and also has much lower costs. In addition, sodium bisulfite is already commonly added to food and drinks, such as wine, fruit juices, jams and dried fruit, where it acts as an enzymatic degradation inhibitor, an antioxidant and bacteriostatic agent. More light needs be shed on the mechanism of action of sodium bisulfite on the AFB<sub>1</sub> molecule. The main reaction product has been isolated and identified as a sulfonate, called 15α-sodium sulfonate or aflatoxin B<sub>1</sub>S (AFB<sub>1</sub>S) (28,29), which forms by insertion of NaHSO<sub>3</sub> at the double bond of furofuranic ring, depriving the AFB<sub>1</sub> molecule of main DNA molecule reaction site, thus reducing mutagenic potential.

### Treatment with Calcium Hydroxide

It has been demonstrated that calcium hydroxide is able to reduce contamination levels when used alone (28) or in combination with formaldehyde. A test on peanut meal (31) showed that this association enhanced  $AFB_1$  detoxification efficacy. The elevated efficaciousness of calcium hydroxide combined with formaldehyde has been verified by in vitro tests (32) and on dairy cows (33). Satisfactory results have also been observed in association with monomethylamine (34). Some authors (35,36) have studied the efficacy of nixtamalization, a traditional practice widely used by South American populations to prepare typical corn tortillas consisting of cooking the corn in boiling water supplemented with calcium hydroxide. This type of treatment achieved corn meal detoxification even if it did not reach safe levels for human consumption. Higher doses of calcium hydroxide reduced the typical organoleptic characteristics but furnished significant results concerning  $G_1$  (AFG<sub>1</sub>) and  $G_2$  (AFG<sub>2</sub>) aflatoxins, whereas  $B_2$  aflatoxin (AFB<sub>2</sub>) and particularly AFB<sub>1</sub> were more stable. The validity of cooking in calcium hydroxide has been verified at rising doses (37) which revealed proportionally increasing levels of AFB<sub>1</sub> destruction (up to 46%). However, the studies also showed the reversibility of the reaction in acid environments and the

greater mutagenic potential of the reaction product compared with the original molecule. Only hazy data are available on the chemical modalities of the molecule's neutralization reaction. Although treatments with sodium, potassium or calcium hydroxides determine only slightly lower detoxification percentages than achieved with ammonia treatments, they present the following advantages:

- 1. Low cost. Calcium hydroxide is the cheapest alkali and is listed in the mineral and chemico-industrial products that may be used in animal feeds.
- 2. Easy application. It is a powder and so can be readily mixed with the feed to be detoxified. Furthermore, it does not require specially built plants using sophisticated technology.

# Treatment with Formaldehyde

Formaldehyde is a compound which is moderately efficacious in attacking and neutralizing the AFB<sub>1</sub> molecule, even if no data on its reactions mechanism are available. Studies showed its enhanced efficacy in association with ammonia (3) and calcium hydroxide (31). In contaminated milk samples addition of 0.5% formaldehyde reduced 1.1  $\mu$ g AFM<sub>1</sub> to 0.05  $\mu$ g (38).

### Other Treatments

Other substances that seem active against aflatoxin include some oxidants, such as sodium hypochrolite, potassium permanganate, hydrogen peroxide and sodium borate. No important studies have been conducted on these treatments.

# Addition of Sorbents

One of the most important approaches aimed at reducing the risk of aflatoxicosis or in limiting decrease in animal performance and toxic metabolite carry-over in milk, meat and eggs, is the use of clays in contaminated feeds to reduce aflatoxin absorption in the intestine. Some in vitro tests (39) showed that various absorbing materials classifiable as aluminas, silicas and aluminosilicates are capable of binding aflatoxin in solution. Extraction using various solvents at different temperatures and pH showed a release which varied in intensity in function of the type of material used. The same authors observed that hydrated sodium calcium aluminosilicates (HSCAS) were particularly efficacious in binding aflatoxin. Definitive information on the molecule AFB<sub>1</sub> fixing modalities cannot be furnished yet, even if chemico-physical adsorption phenomena may occur. The AFB<sub>1</sub> molecule may easily remain imprisoned within the typical complex stratified-reticular structure of the HSCAS. However, the stability of this adsorption in particular pH conditions, such as those in the stomach, should be verified.

Nevertheless, numerous experimental studies have demonstrated the validity of this binding action. Trials conducted on chickens, turkeys and swine fed AFB<sub>1</sub> contaminated feeds and added HSCAS revealed the absence or reduction of typical intoxication phenomena, e.g. increase of the relative weight of vital organs (40,41,42, 43,44,45,46), decrease in body weight or reduced body weight gains and feed conversion (39,41,42,47,48,49,50), brittle bones (51) and metabolite deposition in tissues (52). Some authors (53) obtained successful results by adding HSCAS to contaminated growing lambs' feed. The same authors (54) conducted a detoxification test by adding

HSCAS to dairy cows' feeds characterized by AFB<sub>1</sub> micro contamination level (200 µg AFB<sub>1</sub>/Kg) obtaining a significant reduction in the carry-over and confirming the efficacy of the treatment even at the low contamination levels frequently found in practice.

Analogous detoxification trials have been performed using zeolites, bentonites and modified phylloaluminosilicates. A micronized zeolite (55) was tested as an aflatoxin sorbent in feeds for weaning piglets and it induced a marked reduction in mortality rate and increase of feed consumption and body weight. In contrast, a study on dairy cows (56) did not detect any zeolite induced reducing action on carry-over, while a test on broilers of domestic fowl (57) showed the total absence of beneficial effects determined by addition of zeolite.

These contrasting data can be ascribed to the different experimental methods and, above all, to the type of zeolite used. In fact, in synthetic zeolites, as opposed to natural ones, the pore size distribution varies very little and is generally concentrated within a narrow diameter range. If the size of the pores is compatible with those of the aflatoxin molecules (Table 1), conspicuous adsorption occurs. Vice versa, adsorption can be easily nil because no intermediate sized pores are present.

TABLE 1

Molecular	Sizes	of	Aflato	vine	(58)
Moleculai	SIZES	O1	Allan	יפונגע	201.

Aflatoxin	Molecular Size (Å)		
B <sub>1</sub>	5,18		
$B_2$	5,18		
$G_1$	6,50		
$G_2$	6,50		

A test on bentonite as an aflatoxin sorbent conducted on dairy cows (59) revealed a 33% carry-over reduction, while in vitro trials on trout feed (60) achieved adsorption of 70% the AFB<sub>1</sub> present in the feed. An in vitro test (61) demonstrated the efficacy of a commercial product (Mycobond) made of chemically modified phylloaluminesilicate combined with multilayered montmorillonite and detected the formation of an inert, stable complex capable of preventing absorption of mycotoxins in the intestine.

#### **CONCLUSIONS**

This review furnishes the following conclusions:

Although the main chemical detoxification methods are very efficacious, they do not seem able to fulfill all the requirements, especially those concerning the safety of reaction products and the safeguarding of the nutritional characteristics of the treated foods and feeds. In addition, chemical treatments determine substantial increase in costs which often cannot be met by the breeders.

Addition of sorbents is a simple low cost effective detoxification method which seems to offer good prospectives, even if the vast amount of data available and the contrasting results have not clarified the mechanism of action, nor the materials that can guarantee better results. Therefore, more detailed studies should be performed to detect any possible reactions between the structural characteristics of the sorbents and their adsorption capacity in order to select the most efficacious.

## **ACKNOWLEDGMENT**

We thank Mrs. Moira Macpherson for help with the translation of the paper.

## **REFERENCES**

- 1. FAO-WHO-UNEP. Global perspective on mycotoxins. MYC Document of 4th Conference on Mycotoxins. 1977. Nairobi, Kenya, 19-27 September.
- Council for Agricultural Science and Technology (C.A.S.T.). Aflatoxins and other mycotoxins: an agricultural perspective. 1979. Rep n. 80, 250 Memorial Union, Ames, Iowa 50011.
- Frayssinet, C, LaFarge, C. Process for removing aflatoxins from peanut cake. French Patent 2.098,711, Recorded July 24,1970. Accorded February 14,1972.
- 4. Viroben G, Fremy JM, Delort-Laval J. Traitment à froid des torteaux d'arachide par une solution aqueuse d'ammoniaque: consèquence sur la reduction de la teneur du lait en aflatoxin M<sub>1</sub>. Lait 1983; 63:171-9.
- 5. Fremy JM, Quillardet P. The "carry over" of aflatoxin into milk of cows fed ammoniated rations: use of an HPLC method and a genotoxicity test for determining milk safety. Food Addit Contam 1985; 2:201-7.
- 6. Lee LS, Stanley JB, Cucullu AF, Pons WA Jr., Goldblatt L.. Ammoniation of aflatoxin B<sub>1</sub>. Isolation and identification of the major reaction product. J Assoc Off Anal Chem 1974; 57:626-9.
- 7. Kiermeier, F, Ruffer, L. Veranderung von aflatoxin B<sub>1</sub> in alkalischer losung. Z Lebensm Unters Forsch 1974; 155:129-41.
- 8. Cucullu AF, Lee LS, Pons WA Jr., Stanley JB. Ammoniation of aflatoxin B<sub>1</sub>: isolation of a product with molecular weight 206. J Agric Food Chem 1976; 24:408-10.
- 9. Park, DL, Lee LS, Price RL, Pohland AE. Review of the decontamination of aflatoxins by ammoniation: current status and regulation. J Assoc Off Anal Chem 1988; 71:685-703.
- 10. Lee LS, Dunn JJ, De Lucca AJ, Ciegler A. Experimentia 1981; 37:16-9.
- Lawlor TE, Hawort SR, Zeiger E, Park DL, Lee LS. Mutagenic potential of ammoniarelated aflatoxin reaction products in cottonseed meal. J Am Oil Chem Soc 1985; 62:1136-8.

- 12. Schroeder T, Zweifel U, Sagelsdorff P, Friederich U, Luthy J, Schlatter C. Ammoniation of aflatoxin-containing corn: distribution, in vivo covalent deoxyribonucleic acid binding, and mutagenicity of reaction products. J Agric Food Chem 1985; 33:311-6.
- 13. Haworth SR, Lawlor TE, Zeiger E, Park DL. Mutagenic potential of ammonia-related aflatoxin reaction products in a model system. J Am Oil Chem Soc 1989; 66:102-4.
- 14. Purchase IFH. Acute toxicity of aflatoxins M<sub>1</sub> and M<sub>2</sub> in one-day-old ducklings. Food Cosmet Toxicol 1967; 5:339-42.
- 15. Pong RS, Wogan GN. Toxicity and biochemical and fine structural effects of synthetic aflatoxin M<sub>1</sub> and B<sub>1</sub> in rat liver. J Nat Cancer Inst 1971; 47:585-90.
- 16. Sinnhuber RO, Lee DJ, Wales JK, Landersw MK, Keyl AC. Aflatoxin M<sub>1</sub>, a potent liver carcinogen for rainbow trout. Fed Proc 1970; 29:568.
- 17. Barnes JM. Aflatoxin as health hazard. J Appl Bacteriol 1970; 33:285-98.
- 18. Jothimahalingam R, Govindan S, Punniamurthy N, Balachandran C, Nagarajan R. Effect of feeding aflatoxin-contaminated diet on ammonia treatment in broiler chicken. Indian J Anim Sci 1989; 59:901-2.
- 19. Mahalingam RJ, Govindam S, Punniamurthy, N, Balachandran C. A study on aflatoxin detoxification by aqua-ammonia method in poultry feed. Indian Vet J 1990; 67:149-51.
- 20. Sinha RRP, Arora SP. Biochemical and histopathological changes in liver of chicks fed on aflatoxin containing diets after detoxification. Indian J Anim Nutr 1987; 4:12-9.
- 21. Sharma KS, Chawla JS, Ichhponani JS. Studies on the improvement in the nutritional value of fungal infested groundnut cake. Indian J Poult Sci 1989; 24:179-85.
- 22. Park JM, Son SW, Jean YH, Cho JH, Rhee CJ, Cho TH, Namgoong S, Park KS. Detoxification of aflatoxin in corn using ammonia and yeast. Res Rep Rural Develop Adm Vet 1989; 3:49-53.
- 23. Manson MM, Neal GE. The effect of feeding aflatoxin-contaminated groundnut meal, with or without ammoniation, on the development of gamma-glutamyl transferase positive lesions in the liver of Fisher 344 rats. Food Addit Contam 1987; 4:141-7.
- 24. Fremy JM, Gaultier JP, Herry MP, Terrier C, Calet C. Effect of ammoniation on the "carry-over" of aflatoxins into bovine milk. Food Addit Contam 1988; 5:39-44.
- 25. Moerck KE, McElfresh P, Wohlman A, Hilton BW. Aflatoxin destruction in corn using sodium bisulfite, sodium hydroxide and aqueous ammonia. J Food Prot 1980; 43:571-4.
- 26. Sommartya T, Jatumanusiri T, Konjing C, Maccormac C. Aspergillus flavus in peanut in Thailand with special reference to aflatoxin contamination, and detoxification. Proc Jap Assoc Mycotox, 1988; Suppl. N° 1, 71-2.

- 27. Hagler WM, Hutchins JE, Hamilton PB. Destruction of aflatoxin in corn B<sub>1</sub> with sodiumbisulfite. J Food Prot 1982; 45:1287-91.
- 28. Hagler WM, Hutchins JE, Hamilton PB. Destruction of aflatoxin in corn B<sub>1</sub> with sodiumbisulfite: isolation of the major product aflatoxin B<sub>1</sub>S. J Food Prot 1983; 46:295-300.
- 29. Yagen B, Hutchins JE, Hagler WM, Hamilton PB. Aflatoxin B<sub>1</sub>S: revised structure for the sodium sulfonate formed by destruction of aflatoxin B<sub>1</sub> with sodiumbisulfite. J Food Prot 1989; 52:574-7.
- 30. Dvorak M. Chemical detoxification of aflatoxins. Veterinarni Medicina 1990; 35:37-42.
- 31. Codifer LP Jr, Mann GE, Dollear FG. Aflatoxin inactivation: treatment of peanut meal with formaldehyde and calcium hidroxide. J Am Oil Chem Soc 1976; 53:204-6.
- 32. Piva G, Pietri A, Carini E. Decontaminazione di farina di estrazione di arachide contenente aflatossine mediante trattamento con idrossido di calcio e paraformaldeide. Ann Fac Agric, Catholic Univ Sacred Hearth (Piacenza), 1985; 25:213-22.
- 33. Piva G, Pietri A, Carini E. Detossificazione di farina di estrazione di arachide contaminata da aflatossina B<sub>1</sub> con idrossido di calcio e paraformaldeide e riflessi sul tenore in aflatossina M<sub>1</sub>. Zoot Nutr Anim 1985; 11:303-11.
- 34. Park DL, Jemmali M, Frayssinet C, Lafarge-Frayssinet C, Yvon M. Decontamination of aflatoxin-contamined peanut meal, using monomethylamine: Ca(OH<sub>2</sub>) method. Proc Int Symp Mycotox 1983; 257-66.
- 35. Arriola MC de, Porres E de, Cabrera S de, Zepeda M de, Rolz C. Aflatoxin and tortilla preparation in Guatemala. Aflatoxin in maize. A proceedings of the workshop, El Batan, Mexico, April 7-11, 1986. Ed. by Zuber MS, Lillehoj EB, Renfro BL. 1987; 287-307.
- 36. Arriola MC de, Porres E de, Cabrera S de, Zepeda M de, Rolz C. Aflatoxin fate during alkaline cooking of corn for tortilla preparation. J Agr Food Chem 1988; 36:530-3.
- 37. Price RL, Jorgensen KV. Effects of processing on aflatoxin levels and on mutagenic potential of tortillas made from naturally contamined corn. J Food Sci 1985; 50:347-9.
- 38. Heimbecher SK, Jorgensen KV, Price RL. Interactive effects of duration of storage and addition of formaldheyde on levels of aflatoxin M<sub>1</sub> in milk. J Assoc Offic Anal Chem 1988; 71:285-7.
- 39. Phillips TD, Kubena LF, Harvey RB, Taylor DR, Heidelbaugh ND. Hydrated sodium calcium aluminosilicate: a high affinity sorbent for aflatoxin. Poult Sci 1988; 67:243-7.
- 40. Kubena LF, Harvey RB, Phillips TD, Heildelbough ND. Novel approach to the preventive management of aflatoxicosis in poultry. Proc US Anim Health Assoc 1987; 91:302-4.

- 41. Kubena LF, Harvey RB, Huff WE, Corrier DE, Phillips TD. Efficacy of hydrated sodium calcium aluminosilicate to reduce the toxicity of aflatoxin and T-2 toxin. Poult Sci 1990; 69:1078-86.
- 42. Kubena LF, Harvey RB, Corrier DE, Phillips TD, Huff WE. Diminution of aflatoxicosis in growing chickens by dietary addition of hydrated sodium calcium aluminosilicate. Poult Sci 1990; 69:727-35.
- 43. Kubena LF, Huff WE, Harvey RB, Yersin AG, Elissalde MH, Witzel, DA. Effect of hydrated sodium calcium aluminosilicate on growing turkey poults during aflatoxicosis. Poult Sci 1991; 70:1823-30.
- 44. Harvey RB, Kubena LF, Phillips TD, Huff WE, Corrier DE. Approaches to the prevention of aflatoxicosis explored. Feedstuffs 1988; 60:11-2.
- 45. Harvey RB, Kubena LF, Phillips TD, Huff WE, Corrier DE. Prevention of aflatoxicosis by addition of hydrated sodium calcium aluminosilicate to the diets of growing barrows. Am J Vet Res 1989; 50:416-20.
- 46. Colvin BM, Sangster LT, Haydon KD, Beaver RW, Wilson DM. Effect of a high affinity aluminosilicate sorbent on prevention of aflatoxicosis in growing pigs. Vet Human Toxicol 1989; 31:46-8.
- 47. Lindemann MD, Blodgett DJ, Schurig GG, Kornegay, ET. Evaluation of potential ameliorators of aflatoxicosis in weaning-growing swine. 1987-88 Virginia Technol Livestock Res Rep 1987; 30-4.
- 48. Lindemann MD, Blodgett DJ, Schurig GG, Kornegay, ET. Evaluation of potential ameliorators of aflatoxicosis in weaning-growing swine. J Anim Sci 1989; 67: Suppl. 2, 36-40.
- 49. Haydon KD, Beaver RW, Wilson DM, Cloven BM, Sangster LT. Efficacy of an aluminosilicate in prevention of aflatoxicosis in swine. Special Publication. Georgia College Agric Experiment Station 1990; 67:42-5.
- 50. Phillips TD, Kubena LF, Harvey RB. Prevention of aflatoxicosis in animals and aflatoxin residues in food of animal origin with hydrated sodium calcium aluminosilicate. Healthy animals, safe foods, healthy man. World Assoc of Vet Food Hygienists, X<sup>th</sup> International Symp, Stockolm 2-7 July 1989. Proc 1989; 103-8.
- 51. Chung TK, Baker DH. Phosphorus utilisation in chicks fed hydrated sodium calcium aluminosilicate. J Anim Sci 1990; 68:1992-8.
- 52. Beaver RW, Wilson DM, James MA, Haydon KD, Cloven BM, Sangster LT, Pikul AH, Groopman JD. Distribution of aflatoxicosis in tissues of growing pigs fed an aflatoxin contaminated diet amended with a high affinity aluminosilicate sorbent. Vet Human Toxicol 1990; 32:16-8.

- 53. Harvey RB, Kubena LF, Phillips TD, Corrier DE, Elissalde MH, Huff WE. Diminution of aflatoxin toxicity to growing lambs by dictary supplementation of hydrated sodium calcium aluminosilicate. Am J Vet Res 1991; 52:152-6.
- 54. Harvey RB, Phillips TD, Ellis JA, Kubena LF, Huff WE, Petersen HD. Effect on aflatoxin M<sub>1</sub> residues in milk by addition of hydrated sodium calcium aluminosilicate to aflatoxin contaminated diets of dairy cows. Am J Vet Res 1991; 52:1556-9.
- 55. Stankov M, Obradovic V, Obradovic J, Vukicevic O. Effect of a micronized zeolite preparation on the health and production of weaned piglets. Veterinarsky Glasnik 1992; 46:91-6.
- 56. Pietri A, Fusconi G, Blasi P, Moschini M. Azione sequestrante di una zeolite nei confronti di radionuclidi e aflatossina in diete per ruminanti e monogastrici. Proc X Congresso ASPA, 141-6, Italy, Bologna, 31 maggio-6 giugno 1993.
- 57. Sova Z, Pohunkova H, Reisnerova H, Slamova A, Haisl K. Haematological and histological response to the diet containing aflatoxin B<sub>1</sub> and zeolite in broilers of domestic fowl. Acta Vet Brno 1991; 60:31-40.
- 58. Hesseltine CW. Mycotoxins 1. Mycopathol Mycol Appl 1969; 39:371-83.
- 59. Veldman A. Effect of sorbentia on carry-over of aflatoxin from cow feed to milk. Milchwissenschaft 1992: 47:777-80.
- 60. Winfree RA, Allred A. Bentonite reduces measurable aflatoxin B<sub>1</sub> in fish feed. Progressive Fish-Culturist 1992; 54:157-62.
- 61. Sison JA. Sequestering mycotoxins in animal feeds. Feeds Comp 1992; 12:34-35.

Accepted for publication November 29, 1994.